



MGM
Medical College
Hospital,
Kamothe

INFECTION CONTROL MANUAL

Doc No. MGMH/KAM/QD/HIC 1

Effective Date:
01/06/2021

Revision No: 03


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01/ 01/2022

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HOSPITAL INFECTION CONTROL MANUAL

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AMENDMENT SHEET



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Sr.No.	Section no & Page no	Details of Amendment	Reasons	Signature of the Preparatory Authority
1	HIC 1	Revision of Hospital Infection Control Committee	As per 5 th edition of NABH	
2	HIC 7	Revision of CSSD Policy for reusable devices	As per the Hospital Policy decision	
3	HIC 5	Revision of Hospital Acquired Infection Assessment tool	As per 5 th edition of NABH	
4	HIC 3	Revision of Infection Control Daily round observation sheet	As per 5 th edition of NABH	
5	HIC 6	Infection Prevention and Control guidelines for COVID 19	In view of COVID Pandemic	
6	HIC 6	Revision of audit tool for surveillance activities in high risk areas	As per 5 th edition of NABH	

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I. INTRODUCTION

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Hospital infections are an important cause of morbidity and mortality in the current day scenario. They significantly prolong hospital stay and add to health care expenses. The most effective way to battle them is to prevent them.

Atm.G.m hospital, we have developed a comprehensive infection control program. The policies and procedures that we have adopted are evidence based and updated. They are commensurate with CDC (center for disease control), WHO (World health organization) infection control guidelines and have been adapted to local **needs and National Infection Control Guidelines 2017 draft version by NCDC (National Centre for Disease Control Government of India, Ministry of Health and Family Welfare.**

This manual is an attempt to collate the various infection control policies and procedures at our hospital including those on environment cleaning, hand hygiene, prevention of catheter related infection/healthcare pneumonia/urinary tract infections/surgical site infections, isolation precautions, reporting of communicable diseases, management of occupational exposures to blood and body fluids as well as surveillance of Nosocomial infections. The hospital antibiotic policy is also alluded to as an attempt to prevent antibiotic misuse. To take the antibiotic policy from Dr Urekar 2016, 2017

This manual should educate health care staff atm.G.M. Hospital, Kamothe and further improve infection control practices.
Hospital infection control manual will be revised as and whenever required.

The hospital will develop an annual infection control programme including awareness training and measures to be taken to control and develop the Infection Control practices in the hospital. Plan for the year is attached herewith as Annexure. Committee meetings, Surveillance rounds, training schedule of all categories, Infection Control Week, etc

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Infection Control Committee:

Chairman Medical Superintendent


Team Microbiologist
 Infection control
 Nurse Chief Of Quality

Members Internal Medicine
 Surgeon
 Nursingsuperintenden
 t
 Intensivis
 t

Representativesfrom
 Engineering& Maintenance
 Housekeeping
 CSSD In charge
 OT In charge
 Dietician
 Human Resource
 Administrator
 Blood Bank
 Laboratory
 Imaging center

Functions of Infection Control Committee:

- Reviewand approve a yearly programme ofactivity for surveillance and prevention
- Toreview the surveillancedata and identify areas for intervention. Ensure appropriate stafftraining in infection control and safety
- Communicateand coordinate with other committees to ensure a safe and healthy


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environment.

- Set general infection control policy/guidelines and to provide input into specific infection control issues
- All departments to develop and implement infection control procedures
- Prevent and control Nosocomial infection through periodic surveillance
- Nosocomial infection, evaluation and investigation of infection outbreaks
- Educate staff on medical waste segregation and management.
- Management of occupational hazards such as exposures to blood and body fluids and Needle stick injury.

Functions of Infection Control Team:

- Take hospital visits periodically to ensure all the infection control practices are being practiced
- Report any shortcoming noted to the Chairperson, co-ordinate with the chairperson in planning infection control programs and measures.
- Management of proper isolation technique
- Provision of hand washing or alcohol + sterlium / chlorhexidine based hand cleansing


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solutions

- Development of standards for management of proper insertion of and maintenance of medical devices.
- Works as a clinical supervisor by ensuring all the established policies and protocols are practiced e.g. Hand washing procedures, use of hand rub, isolation policies, care of vascular access and urinary catheters, universal precautions, terminal cleaning and disinfection and follow up of exposure to blood and body fluid.
- Works as an investigator along with the infection control committee to track down outbreaks, evaluate the equipments to detect risks leading to infection hazards
- Works as an educator by participating in formal and informal teaching programs for doctors, nurses and other healthcare workers. Attend appropriate courses and workshops
- Works as a researcher in co-coordinating with the other members of the committee
- Report of microbiology about Nosocomial strains e.g. MRSA, ESBL, Surgical site infection surveillance
- In case of open TB, inform to ward in-charge on floor and immediately discharge or transfer the patient to isolation room

II. UNIVERSAL PRECAUTIONS:

- Universal precautions are to be followed by all health care workers for all patients.
- Staff with abrasions and cuts not to attend patients, without use of waterproof Band-Aid on the cuts. Use gloves whenever required.

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- Washhands before and after work.
- Useonly disposable dressings and syringes.
- Allinen of the infected patients to be soaked in 01% sodium hypochlorite solutions and handed over to laundry staff.

Personal Protective Equipment

a. **Gloves:**


Weargloves (clean, non-sterile glovesare adequate) when touching blood, bodyfluids, secretions, excretions, and contaminated items. Put on clean gloves just before touchingmucous membranes and non-intactskin. Change gloves between tasks and procedures on the same patients after contact with material that may contain a high concentration of microorganisms. Remove gloves promptly after use, before touching non contaminated items and environmental surfaces, and before going to otherpatient,and wash hands immediately to avoid transfer of microorganismto other patients or environments.

b. **Mask, Eye Protectionand Face Shield:**

Weara mask and eye protection or face shield toprotect mucous membranes of the eyes, nose, and mouth during procedures and patient-care activities that arelikely to generate splashes or sprays of blood, body fluids, secretions, and excretions.

c. **Gowns:**

Weara gown (a clean, non-sterile gown is adequate) to protect skin and to prevent soiling of clothing during procedures and patient-care activities that are likely to generate splashes of blood, body fluids, secretions, or excretions. Select a gown thatisappropriate for the activity and amount offluid likely to be encountered. Remove a soiled gown as promptly as possible and wash hands to avoid transfer of microorganisms to other patients or environments.

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Hand Hygiene

- Handwashing is the single most important procedure for preventing Nosocomial infection as hands are an important route of transmission of infection.
- Handwashing involves both mechanical and chemical action. The running water and friction used in cleaning is the **mechanical** action. The soap will emulsify the fat and lower the surfacetension ofwater to facilitate removal ofthe micro- organisms, dirt and oil. This is the **chemical**action.

Indications for routine hand washing and hand antisepsis:

- Beforehaving direct contact with patients
- Beforedonning sterile gloves.
- Beforeand after any procedure
- Aftercontact with blood and body fluid.
- Aftercontact with inanimate objects(including medical equipment) in the immediate vicinity of the patients.
- Afterremoving gloves

What to Use:

- Whenhands are visibly dirty or contaminated wash hands with soap and water
- Ifhands are not visibly soiled, health care worker may use an alcohol-based hand rub.
- Beforeeating and after using of restroomwash hands with soap and water.
- Before giving feed to patient.

Recommendedhand washing agents:

- Liquidsoap
- Alcoholic+ chlorhexidine /sterlium hand rubs
(Hand Washing Procedure- refer to Pictorial Chart)

III. ENVIRONMENTAL CLEANING

Effectiveenvironment cleaning is essentialsince contaminated surfaces may act as a reservoir of potential pathogens. The transfer of microorganisms fromenvironment



to patients is possible through air, water & via handcontact with the contaminated surfaces.

Hospitalenvironment is divided into two Zones:


- **Highriskzone:** - All critical units (MICU,SICU, PICU, CCU, CVTSICU, EMSICU, OT,), Laboratory, Triage, Dialysis, unit, Procedure room, Blood Bank.
- **Lowriskzone:** - Wards, Corridors, Non patientblock, Administrative block
Hotwater and detergent are sufficient for most purposes. Cleaning removes organic matter, salts & visible soils, all of which interfere with microbial inactivation
Thephysical action of scrubbing with detergents and surfactants and rinsing with water during environmental cleaningeffectively removesmicroorganisms.

Cleaning schedule:

Area	Cleaningtime
Lowriskzone	Ineachshift once and whenever required.
Highriskzone	Ineach shift once and whenever required
Walls-General area	Twicein a week and whenever required
Walls- Patient area	Beforeand after patient use. And once every night
Ceilingand Fan	7 daysand after discharge
Lobby	Ineach shift and whenever required

Discharge cleaning:

- Ondischarge of a normal patient, clear all furniture fromthe room.
- Remove the bed linen
- Changecurtains.
- Dodry dusting.
- Cleanall furniture, bed, mattress, and bedside locker withapproved disinfectant solution.
- Washbathroomwith detergent and water.
- Cleanfan, AC ducts, exhaust fan, walls.
- Mopthe floor with detergent and water.

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Cleaning schedule of OT:

Before surgery:

- All horizontal surfaces within the OT are damp dusted before the first scheduled surgical procedure of the day with a clean lint free cloth moistened in the approved disinfectant solution.
- Visual inspection of commencement of the first surgical case.

During surgical procedure:

- Accidental spillage in the area outside the surgical field should be promptly cleaned by placing tissue papers over it then pouring 1 % sodium hypochlorite over it.
- Leave it for 10 minutes then collect it in the scoop, then mop with a disinfectant.
- Discard the contaminated disposable items in yellow bag.
- Then mop the floor with disinfectant solution

In between surgical procedure:

- Conduct a visible check to inspect cleanliness of the operation theatre
- Reusable suction bottles are emptied and cleaned under running water and dipped in sodium hypochlorite for 20 min rinse with water and replace.
- Floor, OT table, OT light and other equipments cleaning to be done with disinfectant solution.

End of the day:

- Terminal cleaning to be done with approved disinfectant.
- All furniture, wall surface, fixed and ceiling mounted equipments; anesthetic equipments should be cleaned with disinfectant.
- Accessories, handles of cabinet are to be disinfected.
- Scrub to be cleaned with detergent solution under running tap water.
- Floor cleaning to be done with approved disinfectant.
- Bathrooms and toilets to be cleaned with detergent powder & Harpic
- Suction bottles to be emptied, cleaned and disinfected by immersing into 1%



sodium hypochlorite solution for 20 min, in case infected for 1 hour.

- Transport vehicles, including straps and attachments and instrument cabinet to be cleaned with approved disinfectant.

Weekly cleaning (Performed on Sunday):

- Remove all movable equipments and furniture from the OT
- Wash with soap, water & disinfectant solution
- A.H.U. to be cleaned with dry vacuum cleaner (ducts and filters to be cleaned weekly and changed as required)
- Ceiling and walls to be cleaned with dry vacuum cleaner
- Floor cleaning to be done with approved disinfectant
- Fogging to be done at night

Periodical cleaning (Done every 6 months):

- It is a 2 day programme
- The ceiling area is opened and cleaned with dry vacuum and sprayed with disinfectant solution
- Ducts to be cleaned
- Ceiling is to be reestablished
- Floor cleaning is to be done with approved disinfectant solution
- Walls and ceiling are sprayed with disinfectant solution
- Fogging to be done.

Surveillance:

- Air sample & swab to be taken weekly basis, every Sunday morning and proper records to be maintained with OT head nurse.

Scrub suit policy:

- There are 4 color coding of scrub suit



- Bluecolor: for doctors
- Greencolor: for Nurse
- Marooncolor: for technician
- Greycolor: for GDA's

Oneshould not enter the O.T without changing into scrub suit.

Noone should leave the O.T wearing O.T clothes and slippers in any situation.

Instrumentcleaning & de-contamination:

- All used instrument to be rinsed with soap &water and then cleaned with disinfectant.....Composition, then wash it with plain water.
- Sendthe instruments to CSSD in closed container for further treatment.
- For expensive, reusable items like endoscopes- refer to manufacturers manual for disinfection & reuse

Out Patient Area &Treatment room:

- Usedisinfectant for floor moping
- Wipe all tabletops, examination table, with disinfectant (0.5%) Composition lint free duster
- Wipe dressing trolley with spirit
- Changeall curtains every 15 days
- Changelinen on examination tableevery day or as and when required

Isolation room/Ward:

- Changecurtains every 7 days and after discharge of one patient
- Airductsto becleanedperiodically (once a week)
- Foggingto be done with approved disinfectant solutionafterdischarge of infected patient
- Cleaning with approved disinfectant solution to be done in each shift
- Admit a patient only after 1-2 hours after fogging
- Bathroom & Toilets should be cleaned with Phenol, twice in a shift

Disinfection of Blood and Body Fluids Spillage: Add the Spill Management Protocol



- Spillage of Blood/Body fluid over the floor
- Soak it with dry absorbent cotton/ tissue and discard it in the yellow bag
- Cover the residual spillage area with 1% sodium hypochlorite solution for 20 minutes
- And then mop it
- Proceed with normal cleaning.

Important Aspects to remember while cleaning:

- Do not flick the dust while dusting or dry mopping
- Change curtains once every fortnight
- Avoid using the patient's linen for dusting.
- Put 'cleaning' sign board or 'wet' signage or 'spillage' signage
- Avoid cleaning in visiting hours.

IV. DISINFECTION AND STERILIZATION:

Medical and surgical devices may serve as vehicles for the transmission of infectious disease to susceptible host. Therefore it is most important that all health care facilities should have comprehensive disinfection policy. The aim of disinfection policy is to make items and equipments safe for patients use by effectively removing microorganisms by cleaning, disinfection and sterilization.

Cleaning:

Cleaning of instruments before decontamination is an essential procedure. This allows the physical removal of dirt and microorganisms. It prevents inactivation of the disinfectants by organic matter and allows complete surface contact during further decontamination procedures

Disinfection:

- It is a process that inactivates non-spore forming infectious agents, using either thermal or (moist or dry heat) or chemical means
- Chemical disinfection does not necessarily kill all microorganisms present but reduce them to a level not harmful to health. The level of disinfection



depends upon the category of the instruments.

Antiseptics:

- Chemical used to kill microorganisms on skin or living tissues are known as antiseptics.

Sterilization:

- It is a process, which achieves the complete destruction or removal of all microorganisms, including bacterial spore.

Categories of patient care equipment:

Classification	Item use	Goal	Appropriate process
Critical items	Items entering sterile tissue, the body cavity, the vascular systems and non-intact mucous membranes e.g. Surgical instruments	Objects will be sterile (free of all microorganisms including bacterial spores.)	Sterilization (or use of single use sterile product)
Semi-critical Items	Items that make contact, directly or indirectly, with intact mucous membrane or non-intact skin e.g. Endoscopes, anesthetic	Objects will be free of all microorganisms with the exception of high numbers of bacterial spores	High level disinfection; thermal disinfection; chemical disinfection
Non-critical Items	Objects that come into contact with intact skin but not mucous membranes e.g. Crutches, BP cuffs.	Objects will be Clean	Low level Disinfection Cleaning (manual or mechanical)

Disinfection of patient care utilities:



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Items	Cleaning/disinfection	Time Required	Frequency
Nebulizerset	Soap& water wash, dry & place on the trolley	Whenever found dirty	Individual Preference
Stethoscope	Alcoholswab/ Sterlium	1-2 mins.	Aftereachuse
Thermometer	Soap &water. Then wet swab followed by dry cotton swab	2-3 mins.	Individual for each patient. Cleaftereach use.
Laryngoscope	Blade-withsoapandwater.	30mins.	Eachuse
	Handle& blade -Alcohol	2-3 mins.	
Nasalprongs (no tube)	Soap&water(onlyif dirty)	2-3 mins.	After each use
Oxygenmask	Soap&water& disinfectant	2-3 mins.	After each use
Ambu bag	Soap& water Infected patient- Disinfect by approveddisinfectant.	Dipfor 1 hour	Aftereachuse
Sputummug,Urinal, Bedpan	Soap &water:Immerse in 1%sodiumhypochlorite.	20to 30 mins.	Aftereachuse& once at night
Ventilator parts	Note: Refermanual. 1. Puret&Bennet company two ventilators 840 , 760 Autoclave the inspiratory & expiratory filters 2. E 360 has disposable filters as	NA	Individual Preference
Ventilator	Bacillocid spray or D125 Microgen	NA	Everyday
Emesis basin,Measuringcup	Soap& water	NA	Aftereachuse
Bpcuffs	Cloth-Soap & water Plastic material- Alcohol swab	NA	Wheneverrequired
Multipara Monitor	Spray What kind of spray		
Monitorcables	Alcoholswab	NA	Aftereachpt.&wheneverrequired
Mattress	1% hypo	20 minutes contact	

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Methods for sterilization:

Refercssd SOP Place it in the appendix typed and ready


Important aspects:

- Allused articles should be disinfected in dirty utility room then dryit and send to the CSSD.
- Surgical Instruments:shouldbe sent in clean cabinet.
- Properrecord should be maintainedat all departments and in CSSD.
- Departments should return items nearing expiry date to CSSD one day prior to the date of expiry

V. INFECTION CONTROL POLICIESFORVARIOUSDEPARTMENTS:

Critical Care Area (MICU, SICU,CCU, CVTS ICU, PICU, EMS ICU):

- Before entering footwear to be removed & kept on racks outside the facility; followed by walk through clean area, then put on rubber slippers.
- Strictbarrier nursing care must be given to the patient.
- Entryof visitors should be minimized.
- Thoroughperiodic hand washing and uses of hands rubs
- (4%or 2% chlorhexidine+alcohol) before and after every patient care.
- Afterhand washing, hands tobe dried by tissue paper
- Disposableitems should be used for all patients.
- Aftersingleuse, all disposable items such as suction catheters, nasal prongs, facemask, enema catheters to be discardedas per bio-medical waste management protocols.
- Wearsterile gloves every time while handling the invasive lines(central line, dialysis line, etc.)
- Allthe color coded bags to be placed for proper segregation of the items.
- Punctureproof container (with properlabel) to be placed for sharps.
- Allthe cuts and abrasions of HCW to becovered with water proof Band-Aid while caring the patient
- I/Vcannula to be changed SOS (if any signs and symptoms of inflammation,

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redness, or if IV is out.)

- Ventilator's cleaning
- Use disposable tubing's (Respiratory circuit) for ventilators with 360 and Suma.
- Change the bacterial filters every 48 hrs when used. And the catheter mounts every 24 hrs and/or so.
- Filter to be sent for autoclave for ventilators 840 and 760.
- Distilled water to be changed every 8 hrs in flow meter.
- Disinfect all electrical equipment with Bacillocid spray / D 125.

Infection Control Policies for Endoscopy room:

General instructions:

- Before entering footwear to be removed & kept on racks outside the facility; followed by walk through clean area, then put on rubber slippers.
- All health care workers must wear the protective gown
- Disinfection of the unit should be done with approved disinfectant solutions i.e. D 125 solution.
- All the procedures must be done with aseptic techniques.
- Strict restrictions of the visitors inside the procedure room.
- No sweeping, only dry mopping of the walls and floors to be done.
- Patients should be provided with O.T gown or dress before procedure.
- T.V monitors should be cleaned with clean cloth (Bacillocid + lint free duster)
- Temperature (20°C-25°C) should be maintained in the procedure room. (Get a thermometer and measure temperature) Sis Aswini will maintain the ambient temperature chart
- Thorough hand washing and use of hand rub (4% or 2% chlorhexidine + alcohol) before and after every patient care. Label the area for hand washing and Instrument cleaning

Cleaning and Disinfection of Endoscope:

- Thorough manual cleaning of the instrument and its internal channels is the most important part of the disinfection procedure.
- Cleaning with warm water with an enzymatic cleanser (Dip the scope in 10% rapid enzyme for 5 minutes and flush it properly).



- All the channels to be flushed and brushed, if accessible, to remove all organic materials (e.g. Blood, tissue) and other residue.
- Reusable accessories (e.g. Biopsy forceps or other cutting instrument) that break the mucosal barriers to be cleaned with warm water and enzymatic cleanser and then dip in disinfectant and rinsed with distilled water between each patient.
- All the cleanser solution to be discarded after each use.
- After proper cleaning the entire scope has to be completely immersed in disinfectant for specified time, after that the scopes to be washed with distilled water. **PL SPECIFY THE TIME**
- The instrument and its channels should be thoroughly forced air-dried. A final drying step that includes flushing all channels with alcohol followed by purging the channels with air greatly reduces the possibility of the endoscope contamination by microorganism
- Before the procedures, the scopes to be thoroughly cleaned and flushed with normal saline or distilled water. **Pl verify this step**
- Scopes should be stored in scope container.

Infection control policy in Dialysis:

General instructions:

- Before entering footwear to be removed & kept on racks outside the facility; followed by walk through clean area, then put on rubber slippers.
- Aseptic technique to be maintained
- All health care workers must wear the protective gown
- Strict restriction of the visitors inside the unit.
- No sweeping, only wet mopping of the walls and floors to be done.
- Temperature (20°C-25°C) should be maintained inside the unit. **INDENT A THERMOMETER AND MAINTAIN REGISTER FOR TEMPERATURE MONITORING**
- A separate Dialysis Machine is used for Hepatitis-B positive and Hepatitis-C positive patients.
- For reactive patients the machines to be kept separately. Never dialyze the non-reactive cases with positive cases
- All non-reactive patient's blood samples to be investigated every 3 months interval
- Hepatitis B vaccination must be done for all HCW who will be posted in Dialysis unit
- Hepatitis- B Vaccination of Non- Reactive Patients in Dialysis Unit



- Beforedialysis, pre vaccination screening to be done
- 1ml Hepatitis B vaccine to be administered
- 1st dose any day
- 2nd dose after one month from 1st dose
- 3dose after 6 months from 1st dose
- After1month of administration of 3rddose, Anti hbsag Titer to be checked
- Nextdose will be decided according to the Titer level

Post - Dialysis:

- Careof Tubing
- Rinsewith RO water.
- Fill up with hypochlorite solution5% and keep for 30 minutes.
- Rinseagain with RO water and remove all the clots.
- Filltube with Renaline4% solution and preserve it in respective chambers ofeach patient.

Care of Cleaning Dialysis:.....

- Rinsewith RO water.
- Fillit with Renaline4% solution and preserve accordingly in the chambers of each patient.
- Careof Machine
- Rinsemachine (for 15 minutes with water)
- Rinsewith the chemical (Bleach 10% for15 minutes) (Depends on themachine)
- Waitfor 15 minutes
- Againrinse with waterfor 15 minutes.

Cleaning of unit:

- Cleanpatients bed after each dialysis with approved disinfectant solutions.
- Moppingshould be done in each shift and a required with approved disinfectant solutions.
- EverySaturday end of the day, clean whole unit with warm water and detergent.
- Cleanallfurniture oftheunit with approved disinfectant solutions.
- Cleandressing trolley and injection loading area.
- Spray approved disinfectant solutions, and closes the unit for 1 hour. Then do



through moping of unit.

- Before and after cleaning take culture swab for surveillance and surveillance record to be maintained with head nurse.

Operation theatre:

- Physical design elements
- Temperature: between 20 to 22 degree centigrade.
- Humidity: 50% to 60%.
- Air handling unit: 5 microfilters changes per hour.
- Airflow should be unidirectional, positive airflow.

Laminar Air Flow:

- Airflow is unidirectional.
- Total air changes 20-30/ hour
- Positive air pressure with velocity 110ft/min at filter point and 50-70 ft./min at the operating table level.
- Filters used are pre filter of 10 micron, micro filters of 5 micron and hepa filters of 0.3 microns. Verify with Mr AMOL

Methods of disinfection:

- Surface cleaning
- Fogging
- Keep air conditioning switched off
- Switch off all lights
- All equipments to be covered with plastic
- Keep room closed for 1-2 hours
- Switch on exhaust for 15 minutes prior starting AC
- Air conditioning to be started after 1 hour of the procedure.



Preparation and concentration of disinfectant for fogging:

For hydrogen peroxide and silver nitrate:

Space Cu.Ft.	Dilution	Fogging duration
1000	200ml in 1000ml	20min
2000	400ml in 2000ml	40min
3000	600ml in 3000ml	60min

Infection control policies in pathology:

Refer laboratory SOP

Infection control policies for Laundry:

General instructions:

- Healthcare Workers, involved in collection, sorting, transportation, and washing of soiled linen should be trained
- All the Health Care Workers must wear mask, plastic apron, during the laundry process
- During sorting, all the hcw must wear mask & gloves
- All the infected linen (infected cases such as tuberculosis, diphtheria, skin lesion, blood borne pathogens HIV, HIV+ve and multi drug resistant bacteria) to be disinfected with approved disinfectant solutions in ward itself.
- All the infected wet linen to be squeezed out properly before sending to laundry
- All the blood soaked linen (green linen from OT) to be disinfected with approved disinfectant solution and washed with water before sending to laundry
- If employee's dress is soiled with infected blood or body fluid put in yellow bag and give to laundry
Sis Divya Krishnan – Please check the above



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Laundry Process:

Refer laundry SOP – No policy available

Kitchen:

Refer Kitchen SOP – Policy to be appended

CSSD:


Refer to CSSD SOP – Policy to be appended

Mortuary:

Refer to Mortuary SOP – Sis Richa might be having the Protocol

Blood Bank:

Refer to Blood Bank SOP – Dr Seema , please provide the protocol

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VI. ISOLATION PRECAUTIONS AND POLICY

General instructions:

- Universal precautions are to be followed for all patients admitted to the hospital irrespective of their diagnosis.
- Special transmission based precaution regarding blood borne pathogen isolation (BBP), airborne pathogen (droplet) isolation and contact isolation have to be followed for patients having specific diseases.
- When a patient is admitted, the exact diagnosis may not be known and hence based on symptom complex, the isolation method is provided below.
- Once the diagnosis is established; initiation, change or stoppage of an isolation method may be warranted. The list of disease with isolation method and duration of isolation is also listed.

Definitions:


Airborne isolation:

- Negative-pressure room preferred.
- For patients with chicken-pox, measles only immunized staff to provide care
- For patients with open tuberculosis, wear mask before entering the room
- Limit patients movement, place a surgical mask if patient is to be transported outside

Contact isolation:

- Private room preferred or else cohort patient with patient of similar illness
- Wear gloves prior to any patient contact
- Wear clean non sterile gown if any contact with body fluids, infectious materials or spillage anticipated
- Remove the gloves and gown before leaving the room/patient care area
- Do not touch environment surface, fomites after removing the gloves/gown
- Limit patient transportation.

Droplet isolation:

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- Privateroompreferred, else cohort patient with patient of similar pure illness.
- Weargloves prior to any patient contact.
- Wearclean non sterile gown if any contact with body fluids, infectious materials or if spillageisanticipated.
- Remove the gloves and gown before leaving the room/patient care area.
- Donot touch environment surface,fomites after removing the gloves/gown.
- Limit patient transportation.

Blood borne pathogen isolation:

- Handwashing before and after patient contact
- Weargloves if contact with blood, body fluids, secretions,excretions and contaminated items anticipated
- Wearmask, eye protection and gown before activities likely to generate splashes andsprays
- Takeutmost care to prevent needle stick injury
- Handleall patient care items properly, dispose appropriately waste and sharps.

Empiricisolation precautions to beinstitutedtill diagnosisestablished:



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Sr. No.	Condition	Potential pathogens	Type of isolation
1	Diarrhea	Enteric pathogens	Contact
2	Meningitis	Meningococci	Droplet
3	Petechial rash	Meningococci	Droplet
4	Vesicular Rash	Varicella zoster	Airborne & Contact
5	Maculo popular rash in child	Measles	Airborne
6	Cough, fever and upper lobe Pulmonary infiltrate in an adult	Measles	Airborne
7	Paroxysmal/ severe persistent cough	Pertussis (Whooping cough)	Droplet
8	History of infection or Colonization with multi drug resistant organisms	Resistant bacteria	Contact
9	Skin, wound or urinary tract Infection in a patient with recent stay at hospital or nursing home	Resistant bacteria	Contact
10	Abscess or draining wound that cannot be covered	Staph aureus	Contact
11	Respiratory infection in infants and children	RSV, Para influenza	Contact

Isolation precautions for specific conditions:

Disease	Isolation	Duration
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Abscess(major)	Contact	DI(Duration of Illness)
Celluliteswith drainage	Contact	DI
Chickenpox	Airborne, Contact	Tillall lesions have crusted
Conjunctivitis(viral)	Contact	DI
Diphtheria	Droplet	Untiltwo cultures taken 24 hrs. Apart are Negative
Diarrhea	Contact	DI
Hepatitis a	Contact	For 3years of age duration of hospitalization; In children 3 to 4 years of age, until 2 weeks after onset of symptoms; and in others, until 1 week after onset of symptoms
Hepatitis b	Bloodborne Pathogen	DI
Hepatitis	Bloodborne Pathogen	DI
HIV	Bloodborne Pathogen	DI
Herpes simplex	Contact	DI
Herpes zoster in Immune-compromised or disseminated	Airborne, Contact	Tillall lesions have crusted
Localized herpes Zoster in normal immune status	Contact	DI
Influenza	Droplet	DI
Avian influenza in Humans	Airborne/contaminated	7days (21 days in less than 12 years)
Measles	Airborne	5days after rash
Mumps	Droplet	Tillswellingsubside (usually 9 days)



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Meningococcal Infections	Droplet	24hrs. After starting antibiotics
MRSA	Contact	Tillcultures negative
Pertussis	Droplet	5days after starting antibiotics
H1N1	Droplet/Airborne	Tillfurther investigations
Pneumonia in infants and young:		
Rabies	Contact	DI
Rubella	Droplet	7daysafter onset of rash
Sorethroat/Scarlet Fever	Droplet	24hours of antibiotics
Tuberculosis Pulmonary	Airborne	Till3 sputumsmeas collected on different Daysarenegative

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Policy for Patient with Smear Positive Pulmonary Tuberculosis:

Ascertain whether hospitalization is required for the patient

Hospitalization not required:


- Inform the concerned HOD/consultant for immediate discharge or shifting of the patient
- Meanwhile give the patient surgical mask to wear
- Ensure that there is no post-operative patient or immuno-compromised patient in the vicinity.

A. Hospitalization required:

- ICU care not required.
- Shift the patient to the atb ward.
- Inform all the healthcare workers about the isolation of the patient.
- Keep the door of isolation room closed at all time
- Provide filter mask to the visitors of the patient.
- Restrict the patient's movements to the room itself.
- Once health is stable discharge immediately.

B. Icu care required

- If the patient is on closed circuit ventilation, he/she need not be isolated
- Non-ventilated patients to be kept in negative pressure isolation room
- Patient is step down to a single room
- Institute all air borne precautions in addition

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Policy for Patient with Varicella Zoster (Chicken Pox):

- Shift the patient immediately to the isolation room
- Keep designated immune staff (previous history of chicken pox or those that received two doses of the vaccine)
- Immunize the exposed nurse if not immune
- Keep the door of the room closed at all times
- Institute all air borne precautions and above given precautions till the scabs dry.


Policy for Patient with Methicillin resistant staphylococcus aureus (MRSA): Dr Suhasini to verify the protocol

- Methicillin resistant *staphylococcus aureus* is a human pathogen that is resistant to penicillin, Cephalosporin and a number of other antibiotics.
- Drugs like vancomycin, Teicoplanin and Linezolid are recommended to treat MRSA infection.
- Screen all the patients with any of the following risk factors for MRSA:
 - Interhospital transfer.
 - Admission with central line access in-situ from other hospitals.
 - Admission with Foley's catheter in-situ from other hospitals.
 - Previous surgery or hospitalization during past 12 months
 - Place the patient under contact isolation till report of MRSA status is received

Source of infection:

- MRSA is common in many hospitals. Infected and colonized hospital patients are the major primary reservoirs in the health care setting. Colonization of hospitalized patients depend on:
 - Length of hospital stay
 - Severity of underlying disease.
 - Presence of wound and / or invasive devices
 - Unhygienic environment
 - Infected patients of MRSA
 - Nutritional status of the patient
 - Carrier status of HCWs and surgical team

Mode of transmission:

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The major route of transmission of MRSA within institution is:

- Patient to patient via hands of the health care workers
- Inadequate hand washing facility
- Inadequate knowledge about infection control practices and policies
- Non-compliance of health care workers for hand-washing practices
- Colonized surface and environment are also significant factors


MRSA patient care:

If patient is identified or detected as colonized in the ward/ICU, following steps to be carried out in department (ward/ICU):

- Isolation of patient
- Handwashing with 2% chlorhexidine scrub (clean and dry with tissue paper) after touching blood, body fluid, secretions, excretions and contaminated items.
- Use of personal protective equipment
- Gloves: use of gloves according to need (sterile or unsterile)
- Mask: use of mask where droplet or airborne infection are anticipated. (E.g. while chest pt.)
- Gown: use clean gowns to protect skin and prevent soiling of clothes
- Plastic aprons while doing any procedure or chances of splashing
- Room surface should be mopped with disinfectant solution
- Daily bath or sponge to patient with 2% chlorhexidine at least for week till 3 negative screenings
- Health education to relative regarding the transmission
- Disposal of waste according to policy
- All used linen should be disinfected before sending to laundry.

Treatment of MRSA Carrier:

- Nasal carriage: 1% chlorhexidine paste thrice daily for 15 days
- Mupirocin in a paraffin base should be applied to the anterior nares thrice daily for 5 days.
- Skin carriage: the staphylococcal load on the skin may be reduced by using an antiseptic detergent for skin hair washing for one week. The agent used includes chlorhexidine, povidone iodine and triclosan
- Treatment of other site colonization Hexachlorophene powder can be applied for colonization of axilla and groin (not to be used on broken skin.)
- Serious infection should be treated with Inj. Vancomycin.

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Clearance:

Infected/ colonized patient should be screened weekly. Three negative screening indicates clearance Health care worker with nasal carriage can continue working once on Mupirocin which is effective within 24 hours.

Policy for Patient with Hepatitis-B, Hepatitis-C, and HIV in the Ward:

- Staff should follow strict universal precautions during care of the patient
- Cases diagnosed outside should be confirmed after admission of the patient
- HCW to ensure that all the cuts and the abrasions be covered with waterproof band-aids while caring for this patient
- HCW giving care to the patient must have received completed dose of Hep-B vaccine
- All used linen patient should be disinfected before sending to laundry

Pre-Operative Preparation:

- Ensure that pre-operative profile is done for the patient
- Cleaning of the part should be done with chlorhexidine or 0.5% betadine
- All the sharps should be disposed along with needle in the puncture proof container
- All healthcare Workers should wear gloves while preparing the patient
- Prior intimation to OT

In the Operation Theater:

- Post as last case of the day
- All the team members should wear double gloves, goggles and protective clothing during the procedure
- Disposable patient draping should be used
- All the instruments used for the procedure to be disinfected thoroughly with disinfectant and then plain water
- All the used linen to be dipped in the disinfectant solution for one hour and sent to the laundry
- After shifting the patient, all the surfaces, wall and table to be cleaned with approved disinfection solution and thereafter fogging, after 3-4 hours thoroughly clean all the surfaces with disinfectant solution.



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VII. PREVENTION OF Healthcare Associated Infection

A. Prevention of Catheter Related Blood Stream Infection (CRBSI)


Peripheral catheters:

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- Establish the vein prior disinfection. Upper extremity preferred over lower extremity
- Practice procedural hand washing technique with soap & water OR Use hand rub.
- Disinfect the site selected using "3 swab method" with isopropyl alcohol + Chlorhexidine and wait till dries.
- Clean gloves are appropriate in non-touch technique is used
- Do not touch the site un-gloved after disinfection
- Do not reuse a vascular access device
- Leave site visibly dry after access is established
- Apply dressing
- Change device if any signs of infection, infiltration & phlebitis or patient complaint of severe pain.

Central venous Catheters (CVP) Insertions:

- Sub-clavian preferred over jugular preferred over femoral
- In children no such preference, use the route which is most comfortable
- Use minimum number of lumens
- Antibiotic coated catheters superior to routine catheters if they are expected to remain in place for more than 5 days SIS DIVYA PL CHECK
- Practice surgical hand washing prior to procedure
- Use maximum barrier precautions (cap, mask, sterile gown and gloves.)
- Clean the site with isopropyl alcohol and 2% chlorhexidine or 10% betadine may be used. Clean in circular manner each (1 min) time for 3 times if povidone iodine is used allow at least 2 minutes for drying
- Insert catheter with aseptic precautions
- Leave site dry after insertion
- After insertion clean the area with 2% chlorhexidine and pat dry with sterile gauze and apply the Micropore dressing with plain sterile gauze
- Do not apply any ointment, betadine, Mupirocin
- Dressing and maintenance
- Regular dressing every 2 days for gauze and 7 days for transparent dressing
- Change dressing earlier if damp, loosened or soiled
- Proper hand hygiene with sterile gloves before dressing
- Inspect for any evidence of catheter site infection
- Clean with 2% chlorhexidine in circular manner dry with sterile gauze apply sterile gauze and apply dressing

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- Clean catheter and stopcocks with 2 % chlorhexidine & wrap the 3 ways with sterile green towel.
- If multi lumen catheter is used designate one port exclusively for hyperalimentation.
- Clean all stopcocks with 70% alcohol or 2 % chlorhexidine prior to use.
- Cap all stopcocks when not in use.

Removal:


- Remove when no longer necessary
- No need to send routine surveillance cultures from the catheters. If tip of catheter is send for cultures then send parallel blood cultures from peripheral site along with tip for culture
- Replace catheter if there is any evidence of infection at exit site
- Remove all catheters if person is hemo-dynamically unstable and CRBSI is suspected
- If CRBSI is suspected do not replace catheters over a guide wire.

Arterial catheters:

- These same principles for insertion, maintenance and removal as CVC apply
- Use disposable transducers preferably. Use sterile reusable transducers in accordance with manufacturer's instruction if disposable transducers not available
- Replace the transducer at 72 hours interval along with other components of the system including the tubing, the flush solution and continuous flush device
- Keep all components of the pressure monitoring system sterile
- Minimize manipulations and keep a closed flush system
- When the pressure monitoring system is accessed through a diaphragm rather than stop cock wipe diaphragm with 70% alcohol or 2 % Chlorhexidine prior to access
- Do not use any parenteral fluids or dextrose containing fluids through the system

Umbilical catheters:

- These same principles as for CVC apply
- Umbilical artery catheters should ideally not to be left for more than 5 days. Can be kept up to 2 weeks if aseptic precautions followed.
- Remove earlier and do not replace if CRBSI, thrombosis, vascular insufficiency is suspected.


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Administration sets, fluids, medication:

- Blood and Blood Products: Single Use
- Lipid, Amino acid : 24 hours
- Primary Infusions : 72 hours
- Antibiotics: 72 hours
- Others: 72 hours
- Use collapsible bags for IV fluids whenever possible especially for patients at high risk for Healthcare associated infection (avoid using needles for air inlets.)
- Preferably use single dose vials.
- If multi-dose vials are used, refrigerate after each use and wipe the access surface with 70% alcohol before inserting the needles.

B. Prevention of urinary tract infections (CAUTI):

- Educate personnel for correct techniques of catheter insertion and care. Periodically re-educate HCW in catheter care
- Catheterize only when necessary. Condom catheter drainage, supra-pubic catheterization, and intermittent urethral catheterization can be useful alternatives to indwelling urethral catheterization
- Use smallest suitable bore catheter consistent with good drainage and to minimize urethral trauma
- Perform hand washing with soap & water or use hand rubs
- Insert catheter using aseptic technique and sterile equipment. Use sterile gloves, sterile drape, swab, lubricant jelly and antiseptic solution (betadine). Secure catheter properly to prevent movement and urethral traction
- A sterile, continuously closed drainage system to be maintained
- The catheter and collecting bag should be emptied regularly using a separate collection container for each patient (the draining spigot and nonsterile collecting container should not come in contact.)
- The collecting bag should always be kept below the level of the bladder
- If breaks in aseptic technique, disconnection, or leakage occur, the collection system should be replaced using aseptic technique after disinfecting the catheter-tubing junction
- If small volumes of fresh urine are needed for examination, the distal end of the catheter, or preferably the sampling port if present, should be cleansed with a disinfectant, and urine then aspirated with a sterile needle and syringe

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- Large volumes of urine for special analysis should be obtained aseptically from the drainage bag
- Daily care to be given with betadine solution
- Regular bacteriological monitoring of catheterized patients is not recommended
- Changing for catheter and bag
 - Simple Foleys Cath.: 14 days
 - Silicone Foleys Cath: 3 months
 - Urine collection bag: With Foleys Cath. Or If Spoiled, any leakage

C. Prevention of health care associated pneumonia: Ventilator associated pneumonia (VAP)

- Use noninvasive ventilation to reduce the need or shorten the duration of endotracheal intubation.
- Unless contraindicated by the person's condition perform orotracheal rather than nasotracheal intubation.
- Perform endotracheal intubation with sterile technique after procedural hand wash/hand rub and sterile gloves.
- Avoid frequent endotracheal intubation.
- If feasible, use an endotracheal tube with a dorsal lumen above the endotracheal cuff to allow drainage (by continuous or frequent intermittent suctioning) of tracheal secretion that accumulates in the patient's sub-glottic area.
- In the absence of medical contraindication elevate head end at 30 to 45 degree to prevent aspiration.
- Perform good chest physiotherapy for incubated patients. And do proper suctioning with aseptic precautions.
- Maintain good oral hygiene to prevent oropharyngeal colonization and subsequent aspiration.
- Change the ventilator tubing whenever required and use only single use tubing.
- Routinely verify appropriate placement of the feeding tube.

Prevention of postoperative pneumonia:

- Instruct preoperative patients, especially those at high risk for contracting pneumonia, about taking deep breaths and ambulating as soon as medically indicated in the post-operative period

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- Insist patient for chest physiotherapy (incentive spirometry) on post-operative
- Care of patients with tracheostomy
- Perform tracheostomy under aseptic conditions
- Daily tracheostomy care to be given
- Follow aseptic precautions while doing tracheostomy suction
- When changing a tracheostomy tube, wear a gown, use aseptic technique and replace the tube with sterile new tube.


D. Prevention of surgical site infections (SSI)

Pre-operative:

- Whenever possible, identify and treat all infection remote to the surgical site before elective operation and postpone elective operations on patients with remote site infection until the infection has resolved
- Do not remove hair preoperatively unless the hair or around the incision site will interfere with the operation
- If hair is removed, remove immediately before the operation, preferably with clip removal
- Control blood glucose of diabetic patients
- Require patients to shower or bathe with 4% chlorhexidine twice pre-operative or at least the night before the operative day
- Encourage tobacco cessation in any format least 30 days before the elective operation
- Keep preoperative hospital stay as short as possible.

Peri-operative:

- Keep nails short and do not wear artificial nails. Do not wear hand or arm jewelry
- Change into scrub suit and wear appropriate personal protective equipments (cap, mask, eye protection,)
- Do surgical hand washing
- Do not wear sterile gown and sterile gloves
- Change the PPE when visibly soiled, contaminated, and/or penetrated by blood or other potentially materials
- Use appropriate antiseptic agent for skin preparation of the

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patients(70%alcohol,10%povidine iodine, 4% chlorhexidinegluconate)

- Followthe antibiotic policy for antibiotic prophylaxis
- Handletissue gently, maintain effective hemostasis, and minimize devitalized tissue and foreign bodies (i.e. Sutures, charred tissues, necroticdebris.)And eradicate dead space at the surgical site
- Ifdrainage is necessary, use a closed suction drain. Place a drain thought a separate incision distal fromthe operative incision. Remove the drain as soon as possible

Post-Operative:

- Protectwith a sterile dressing for 24 to 48 hours post-operatively an incision that has been closed primarily
- Washhands before and after dressing and any contact with the surgical site
- Whenan incision dressing mustbe changed, use sterile technique
- Educatethe patient on discharge regarding proper incision care, symptomsof SSI, and the need to report such symptoms
drsuhasini shall take it up and do the needful

Antibiotic prophylaxis for surgery

- Refer antibiotic policy.



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VIII. ANTIBIOTIC POLICY:

- Refer Antibiotic policy.



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IX. BIO-MEDICAL WASTE MANAGEMENT:

The hospital follows 3 color codes for segregating waste and disposal as per guidelines.

Blackbag:

This encompasses general waste & kitchen waste, is sent to the NMMC for final disposal. It includes:

- Office papers,
- Paper cups,
- Tissue papers,
- Kitchen waste.

Yellowbag:

This encompasses the infectious waste which is handed over to Mumbai waste management ltd. It includes:

- Human tissues.
- Organs and body parts.
- Blood and body fluid tinged dressing materials.
- Microbiology waste.
- Pathology waste.

Bluebag:

This encompasses the infectious plastic waste which is handed over to Mumbai waste



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management Ltd. It includes:

- Syringes.
- Catheters.
- All Tubing.
- Ivbottles.
- Cap&mask.
- Teenasheet.
- Adult& baby diaper
- Gloves

Punctureproof container:

This encompasses what all which can puncture skin which is handed over to Mumbai waste management Ltd. It includes:

- Blades
- Needles
- Scalpels
- Broken glasses
- Ampoules

Unbrokenglasses and Plastic bottles:

Collection of unbroken glasses and plastic bottles should be in different blue bag at dirty utility. It is handed over for scrap selling.

Change the SOP to the new one Sis Divya Krishnan



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X. OCCUPATIONAL EXPOSURE TO BLOOD BORNE PATHOGENS:

Management protocol:

- Do not squeeze or suck the area
- Do not panic
- Wash the area with plenty of soap and water immediately
- Report to the casualty and give all details of exposure to CMO about exposure/source and your own immunization status
- Inform to supervisor on duty and infection control nurse about incident
- Follow the following chart for treatment.

Post exposure prophylaxis for HIV:

- Exposure of blood and body fluid to intact skin does not require any PEP.

Determine exposure code:

EC-1= Small volume (e.g. Few drops/contact for short duration)

EC-2= Large volume (e.g. Large volume/major splash/contact for long duration) several minutes or more.

EC-3= Deep puncture/ large volume/ visible blood on device.

Determine source code:

Source sero -Ve = SC 1

Source sero +Ve = SC 2

Source unknown = SC 2 \implies SC 3

Use of following treatment regimen as per EC & SC:



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EC

SC

PEP Regimen

1

1

PEP not required

1

2/3

basicregimen

2

2/3

Expanded Regimen

3

2/3

Basic Regimen& Expanded Regimen

2/3

1

Evaluate Risk Factor of Source*

- Source high risk behavior

Yes

No

If 'No', then No Treatment /If 'YES', Follow Basic Regimen.

Basicregimen:

Tab.Duovir(Lamivudine 150mg+ Zidovudine300mg) tab twice a day for 28 days.

Expandedregimen:

Basic+ Indinavir 800mg thrice a day for 28 days.

Highriskbehavior:

Checkwith the high risk assessment of source patient.

Treatmentto be initiated within 4 hours & not later than 48 hours.

PEP for Hepatitis-B virus:

Immunizationstatus of
staff:

Non-immunized

Yes

No

Immunizationschedulecompleted (3Doses)

Yes

No

Partiallyimmunized (one or doses orinappropriately

Yes

No


Antibodytiter known (> 10 IU)

Yes

No

Pepe determinationfor HBV:

Sr. No	Immunizationof staff	Source	Treatment
1	Fullyimmunized	Hbsag-Veor +Ve	No HB Ig.No vaccine
2	Partiallyimmunized	Hbsag-Ve	Followvaccine aspervaccinationschedule.
3	Notimmunized	Hbsag-Ve	Noaction if antibody titers are

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			Adequate(>10 IU/MI)
4	Partiallyimmunized	Hbsag+Ve	Starthbig&follow Vaccinationas per schedule
5	Notimmunized	Hbsag+Ve	Starthbig&followvaccinationas per schedule

Hbig Dose: 0.07ml/kg IM stat.

If antibody titer is unknown: Advice for a titer checks within 24 hours & initiates Ig as per indication.

Follow the below mentioned schedule for vaccination if staff not immunized

0 dose (1ml) at the time of injury

1st dose (1 ml) after 1 month

2nd dose (1ml) after 6 months

XI. HANDLING OF OUTBREAKS:

Definition:

An increase in the isolation rate of an organism or clustering of clinical case in the same time frame suggests as outbreak.

Factors suggesting an outbreak:

- Laboratory report of a bacteriology specimen grows an alerting organism.
- Two or more patients are found to have an infection attributed to a species not previously documented, particularly if it has occurred after a surgical procedure.
- The clinician or ward/department staff reports multiple infection of a similar nature.

Investigation of an outbreak:

- An outbreak is an infection control emergency; measure should be taken as soon as out-break is suspected
- Begin preliminary evolution and determine a background rate of infection
- Confirm the existence of an outbreak
- Confirm the diagnosis using the microbiological methods
- Create a case definition that may include laboratory and clinical data. Start with a



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broad case definition that can be redefined at a later date

- Develop line listing by identifying and counting cases or exposures. Describe that data in terms of time, place and person. Remember that cases may have been discharged from the health care facilities.
- Take immediate control measures. Determine who is at risk of becoming ill. Look at changes that may have affected the rate of infection, e.g. New staff, new procedures, new laboratory test, and health care workers, patient ratio. Etc.
- Communicate information to relevant personnel.
- Write a coherent report (preliminary and final).
- Summarize investigation and recommendations to the appropriate authorities.
- Implement long term infection control measures for prevention of similar outbreaks.

XII. SURVEILLANCE AND MONITORING:

- Lab records scrutiny.
- Infection control nurse examines lab reports daily and discusses it with microbiologist and consultants whenever required.
- Then visit all patients and collect the history of patients as per infection control point of view.
- Daily visits to all wards and department.
- Infection control nurse has to visit all wards and department daily to examine all records of all clinical infections.

Healthcare Associated Infection rates:

- Surgical site infection (SSI)
- Intravascular catheter infection rates per thousand catheter days.
- Ventilator associated pneumonia rates per thousand ventilator days.
- Urinary tract infection rates per thousand catheter days.

Periodical tests done by infection control committee:

Test done on	Tested for	Frequency
Water	Microbiology	Every month
Air samples: OT Dialysis icus Surfaces wabs	Microbiology	Every week
Air samples:	Microbiology	6 weekly



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Wards,ER, OPD		
F& B: Foodplates	Microbiology	Everymonth
Foodhandlers: Stool, blood test& nasal swabs	Stoolfor routine and Microscopy	Everysix monthly

Auditingdone by Infection control team:


- Centrallineaudit
- Peripheralline audit
- Foley'scatheter
- Handhygiene compliance Audit

XIII. EMPLOYEE WELFARE MEASURES:**Hepatitis-B Vaccinationfor Health Care Workers (HCW)****Immunization / Vaccination:**

Immunization against Hepatitis-B will minimize the risk of Hepatitis-B transmission through blood and body fluid contacts. All the health care workers are immunized against Hepatitis-B vaccination. The ICN coordinates the programme under the guidance of HR and the Infection Control Committee.

Vaccinationschedule:

Sr. No	Duration	Dose
1.	0Month	1 st Dose (1ml I/M in adults)
2.	1Month	2 nd Dose (1ml I/M in adults)
3.	6Month (fromthe1 st dose)	3 rd Dose (1ml I/M in adults)

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XIV. NOTIFICATIONS TO MUNICIPAL CORPORATION:

The following cases are notified to Municipal Corporation .The data is sent by Infection Control Nurse:

Water Borne Diseases:

- Vibrio cholera
- Hepatitis A and E
- Typhoid
- Filariasis
- Dysentery

Vector Borne Diseases:

- Malaria
- Dengue
- Kala-azar
- Japanese Encephalitis
- Chikungunya

Others:

- HIV/ AIDS
- Acute Flaccid Paralysis (AFP)
- Chicken Pox
- Hepatitis B and C
- Tuberculosis
- Leprosy



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- Leptospirosis
- Polio
- Rabies

XV. ANNEXURE:**All formats used for monitoring the infection control practices.**STANDARDIZATION OF DISINFECTION SOLUTION FOR ICU AND WARDS

Disinfection & Skin antisepsis	Standardization
Hand Antisepsis for Invasive procedures	<u>Surgical Scrub with:</u> 4% chlorhexidine Followed by micro shield (2.5% chlorhexidine and 70% ethanol).
	<u>Surgical Scrub OT:</u> 7.5% povidine iodine thrice /4%chlorhexidine scrub thrice followed by micro shield(2.5%chlorhexidine and 70% ethanol)
Skin Preparation	<u>Peripheral line insertion</u> : micro shield, chlorhexidine 2.5% and 70% ethanol
	<u>Blood culture:</u> 10% betadine three times followed by micro shield (2.5% chlorhexidine and 70% ethanol)
	<u>Central line Insertion:</u> Scrub with 4% chlorhexidine in a circular manner thrice followed by cleaning of the area with micro shield (2.5% chlorhexidine and 70% ethanol).
	<u>Skin preparation:</u> Body bath with soap and water and use the triclosan as a moisturizer and keep contact time of 1 minute and rinse body with plain water. Followed by 10% betadine paint on the designated surgery site / all over the body.



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	<p>Foleys catheter insertion: 5% betadine solution for cleaning the external genital area and for the prepuce. In case of female mucus membrane to be cleaned with Savlon solution 1ml in 15 ml water dilution.</p> <p>Skin preparation for surgery OT:Scrub with 7.5% povidine Iodine twice followed by swabbing with 2.5% micro shield hand rub, wait for it to dry then paint the site with 10% betadine.</p>
Floor and surface Cleaning	<p>Fogging: Eco shield (20%) Surface disinfection during fogging - Eco shield (10%) Electrical equipments - Bacillol 25. Disinfectant solution in Sharp containers - Lonza ID-50 (2%). Daily surface disinfection – Bacilloid Extra (1%)</p>
Decontamination of non – critical instruments used for patient care	Solution Lonza guard ID 50 2% (400 ml in 20 liter of water)
Disinfection of equipments used for care of patients	<p>Chittle forceps: whole unit to be autoclaved daily and to be kept dry.</p> <p>Cidex OPA all other critical care equipments that cannot be autoclaved contact time 10 minutes.</p>
Corridors and General lobby Cleaning	R2 solution (2-5%)